

A NEW SAPONIN FROM *DEERINGIA AMARANTHOIDES*

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ABSTRACT.—A new triterpenoid saponin, 3-O-[α -L-rhamnopyranosyl (1 \rightarrow 3)- β -D-glucuronopyranosyl]-28-O-[β -D-xylopyranosyl (1 \rightarrow 2)- β -D-glucopyranosyl]-3 β -hydroxyolean-12-en-28-oate **3** has been isolated together with two known saponins, 3-O-[α -L-rhamnopyranosyl (1 \rightarrow 3)- β -D-glucuronopyranosyl]-3 β -hydroxyolean-12-en-28-oic acid **1** and 3-O-[α -L-rhamnopyranosyl (1 \rightarrow 3)- β -D-glucuronopyranosyl]-28-O-[β -D-glucopyranosyl]-3 β -hydroxyolean-12-en-oate **2**, from the fruits of *Deeringia amarantoides*.

Deeringia amarantoides L. Merrill (Amaranthaceae) grows in the Western Himalayas throughout the area between 1000 and 4000 feet (1). It has been reported to possess anticancer (2) and spermicidal activity (3). A thorough survey of the literature showed that there is no report of any chemical work in this genus. Herein we report a new oleanolic acid-based saponin along with two known saponins.

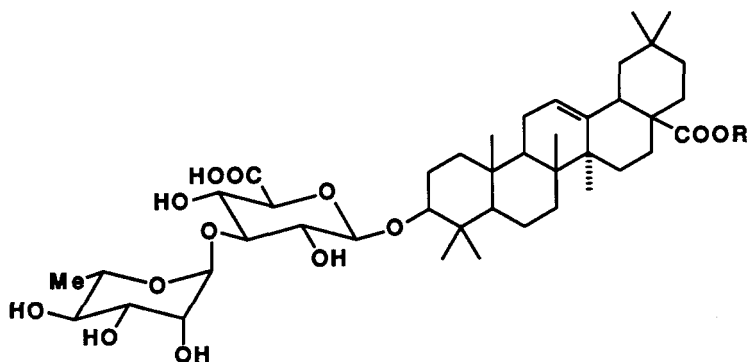
RESULTS AND DISCUSSION

The EtOH extract of the fruits of *D. amarantoides*, on repeated cc, afforded three compounds which exhibited positive color tests for triterpenoid saponins.

Compound **1** in fabms (negative ion) displayed a molecular ion at m/z 777 [$M - H$] $^-$. The peaks at m/z 631 [$M -$

$H - 146$] $^-$ and 455 [$M - H - 146 - 176$] $^-$ arose by the loss of a terminal methylpentose and a methylpentosyl hexose unit, respectively. Compounds **2** and **3** in fabms displayed molecular ions at m/z 939 [$M - H$] $^-$ and 1071 [$M - H$] $^-$ indicating the molecular weights to be 940 and 1072, respectively. Compound **3** exhibited peaks at m/z 939 [$M - H - 132$] $^-$ and 925 [$M - H - 146$] $^-$ by the loss of terminal pentose and methylpentose, respectively. The simultaneous loss of a methylpentose and a pentose unit was concluded from the ion recorded at m/z 793. The peak at m/z 631 was explained by the loss of a methylpentose, a pentose, and a hexose unit from the molecular ion.

Acid hydrolysis of **1-3** furnished oleanolic acid, D-glucuronic acid, and L-

**1** R = H**2** R = β -D-glucopyranosyl**3** R = β -D-xylopyranosyl (1 \rightarrow 2)- β -D-glucopyranosyl

rhamnose; in addition **2** afforded D-glucose and **3** gave D-glucose and D-xylose. Compounds **1–3** were treated with CH_2N_2 and hydrolyzed to afford the methylester of oleanolic acid from **1** and oleanolic acid from **2** and **3**. This confirmed that in compounds **2** and **3** the $-\text{COOH}$ group was not free and glycosylation was at C-3 and C-28. The methylester of compounds **1–3** on NaBH_4 reduction resulted in the conversion of the $-\text{COOMe}$ group to CH_2OH forming the glucosyl derivatives. On permethylation (**4**) followed by hydrolysis, these glucosyl derivatives of **1–3** furnished 2,4,6-tri-*O*-methyl-D-glucose and 2,3,4-tri-*O*-methyl-L-rhamnose, in addition **2** gave 2,3,4,6-tetra-*O*-methyl-D-glucose and **3** gave 2,3,4-tri-*O*-methyl-D-xylose and 3,4,6-tri-*O*-methyl-D-glucose. 3,4,6-Tri-*O*-methyl-D-glucose gave a positive color test with Wallenfel's reagent (**5**) showing a free hydroxyl group at C-2.

Alkaline hydrolysis of **2** and **3** with K_2CO_3 provided a prosapogenol, which was found to be identical to compound **1** (mmp, co-tlc, and co-ir).

The partial acidic hydrolysis of **3** furnished prosapogenols PS_1 , PS_2 , PS_3 , and PS_4 , in addition to oleanolic acid. PS_2 and PS_3 were identical to compounds **1** and **2**, respectively. PS_1 on acidic hydrolysis afforded D-glucose and glucuronic acid, while PS_4 gave D-glucose and D-xylose. Permethyated PS_4 on methanolysis followed by hydrolysis afforded 2,3,4-tri-*O*-methyl-D-xylose and 3,4,6-tri-*O*-methyl-D-glucose.

The types of linkages at the glycosidic bonds were confirmed by the ^1H - and ^{13}C -nmr spectra. In the ^{13}C -nmr spectra of **1**, the C-3 of glucuronic acid was observed at δ 83.9, revealing deshielding of ca. 6 ppm for this carbon resonance in comparison to the reported values for methyl-*O*- β -D-glucoside (**6**) as a result of glycosidation at C-3. The anomeric signals of the D-glucuronic acid and L-rhamnose were observed at δ 5.23 (d, $J = 7$ Hz) and δ 4.98 (s), respectively, in

the ^1H -nmr. Thus, **1** was characterized as 3-*O*-[α -L-rhamnopyranosyl (1 \rightarrow 3)- β -D-glucuronopyranosyl]-3 β -hydroxyolean-12-en-28-oic acid.

^{13}C nmr of **2** indicated that the inner glucuronic acid was substituted at C-3 (δ 83.6 ppm, revealing deshielding of ca. 6 ppm for this carbon resonance) with a terminal rhamnose. ^1H nmr showed anomeric signals at δ 5.79 (d, $J = 8.5$ Hz), 5.22 (d, $J = 6.5$ Hz), and 4.88 (s) showing the presence of β -linked glucuronic acid and D-glucose and α -linked rhamnose. Thus, **2** was characterized as 3-*O*-[α -L-rhamnopyranosyl (1 \rightarrow 3)- β -D-glucuronopyranosyl]-28-*O*-[β -D-glucopyranosyl]-3 β -hydroxyolean-12-en-28-oate. Methyl ethers of **1** and **2** have also been reported for the first time from the aerial part of *Zexmenia buphtbalmiflora* (**8**).

C-3 of glucuronic acid and C-2 of glucose appeared at δ 83.9 and δ 78.1, respectively, in the ^{13}C -nmr of **3**, revealing deshielding ca. 6 ppm and ca. 5 ppm for these carbon resonances in comparison to the reported values (**6**). Anomeric signals in the ^1H nmr appeared at δ 6.32 (d, $J = 7$ Hz), 6.28 (d, $J = 8$ Hz), 4.91 (d, $J = 8$ Hz), and 4.26 (br s). Thus, **3** was characterized as 3-*O*-[α -L-rhamnopyranosyl (1 \rightarrow 3)- β -D-glucuronopyranosyl]-28-*O*-[β -D-xylopyranosyl (1 \rightarrow 2)- β -D-glucopyranosyl]-3 β -hydroxyolean-12-en-28-oate.

EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES.—Mp's were recorded with a Boetius microscopic apparatus, fabms JEOL, JMS-HX-110 instrument in the negative and positive ion mode using MeOH and glycerol as solvent. ^1H -nmr spectra were recorded at 400 MHz using $\text{C}_5\text{D}_5\text{N}$, and ^{13}C -nmr spectra were recorded at 100 MHz using CD_3OD , with TMS as an internal standard. Cc was performed on Si gel 60G (Merck). The spots on tlc were visualized by spraying with 10% aqueous H_2SO_4 followed by heating. Pc was carried out on Whatman paper no. 1 using the descending mode and visualized with aniline hydrogen phthalate. The following chromatographic solvents were used: (A) petroleum ether (60–80°)- Me_2CO (1:1), (B) CHCl_3 -MeOH (60:40), (C) C_6H_6 - Me_2CO (9:1), (D) *n*-BuOH-HOAc- H_2O

(4:1:5), (E) *n*-BuOH-EtOH-H₂O (5:1:4), (F) CHCl₃-MeOH-H₂O (65:40:10), and (G) C₆H₆-EtOAc (8:2).

ISOLATION.—The fruits of *D. amarantoides* were collected from Romdhar (Tehri Garhwal) in July and authenticated by the Department of Botany, University of Garhwal Srinagar. Voucher specimen is preserved in the Ethnobotanical Herbarium of the Department of Botany, University of Garhwal. The air-dried material (4 kg) was defatted with solvent A in a Soxhlet apparatus. The solvent-free mass (3 kg) was exhaustively extracted with 95% EtOH until the extracts became colorless. The concentrated mass was partitioned between *n*-BuOH-H₂O (1:1) (4 × 500 ml). The *n*-BuOH extract, after concentration under reduced pressure, was dissolved in MeOH (50 ml) and precipitated in dry Me₂CO (anhydrous K₂CO₃), to yield a complex mixture of saponins which on repeated cc (solvent B) afforded compounds **1** (10 g), **2** (12 g), and **3** (9 g).

COMPOUND 1.—Colorless needles (10 g) from MeOH: mp 215–220°; ir max (Csl) cm⁻¹ 3400 (OH), 1690; fabms *m/z* 799, 777, 631, 455; ¹H nmr 0.70, 0.72, 0.81, 0.82, 0.84, 0.96, 1.04 (each s, 7 × *tert*-Me group of aglycone), δ 5.16 (1H, br s of H-12 of aglycone), δ 5.23 (1H, d, *J* = 7 Hz), 4.98 (1H, br s) [anomeric protons] ppm; ¹³C nmr (C-1–C-30 of aglycone) 39.7, 28.7, 90.9, 38.9, 57.0, 17.8, 33.4, 40.1, 48.2, 37.8, 24.5, 123.6, 145.1, 42.7, 28.6, 23.9, 47.6, 42.9, 47.5, 31.5, 34.8, 34.0, 27.7, 17.0, 15.9, 17.7, 26.4, 178.07, 34.7, 23.9; rhamnopyranosyl 102.4 (C-1), 72.0 (C-2), 72.3 (C-3), 73.8 (C-4), 69.5 (C-5), 19.2 (C-6); glucuronopyranosyl 106.4 (C-1), 76.2 (C-2), 83.9 (C-3), 72.1 (C-4), 78.1 (C-5), 178.0 (C-6).

COMPOUND 2.—Crystallized from MeOH (12 g): mp 260–265°; ir max (Csl) cm⁻¹ 3400 (OH), 1690; fabms *m/z* 939, 793, 777, 631, 455; ¹H nmr 0.72, 0.82, 0.96, 0.97, 1.01, 1.24, 1.30 (each s, 7 × *tert*-Me group), 5.45 (br s of H-12 of aglycone), 5.79 (1H, d, *J* = 8.5 Hz), 5.22 (d, *J* = 6.5 Hz), 4.88 (s) [anomeric protons] ppm; ¹³C nmr (C-1–C-30 of aglycone) 39.7, 28.6, 91.1, 37.8, 56.9, 17.8, 33.06, 40.0, 47.1, 37.8, 24.4, 123.7, 144.7, 42.8, 28.6, 23.98, 47.9, 42.8, 47.9, 31.4, 34.9, 34.8, 28.7, 17.0, 15.9, 17.7, 26.4, 178.0, 33.9, 23.9; glucuronopyranosyl 106.5 (C-1), 76.2 (C-2), 83.6 (C-3), 72.1 (C-4), 78.1 (C-5), 178.9 (C-6); rhamnopyranosyl 102.6 (C-1), 72.0 (C-2), 72.3 (C-3), 73.8 (C-4), 69.5 (C-5), 19.3 (C-6), glucopyranosyl 95.7 (C-1), 73.8 (C-2), 77.1 (C-3), 71.1 (C-4), 78.1 (C-5), 62.8 (C-6).

COMPOUND 3.—Crystallized from MeOH (9 g): mp 270–274°; ir max (Csl) cm⁻¹ 3400 (OH), 1680; ¹H nmr 0.81, 0.89, 0.92, 0.99, 1.08,

1.26, 1.29 (each s, 7 × *tert*-Me group), 5.11 (1H, br s, H-12 of aglycone), 6.32 (1H, d, *J* = 7 Hz), 6.28 (1H, d, *J* = 8 Hz), 4.91 (d, *J* = 8 Hz), 4.26 (br s) [anomeric protons] ppm; ¹³C nmr (C-1–C-30 of aglycone) 39.8, 28.6, 91.1, 37.8, 56.8, 17.8, 33.0, 40.0, 47.1, 37.8, 24.4, 123.7, 144.7, 42.8, 28.6, 23.98, 47.9, 42.8, 47.9, 31.4, 37.8, 34.8, 28.7, 17.0, 15.9, 17.7, 26.4, 178.0, 33.9, 23.9; glucuronopyranosyl 106.5 (C-1), 72.5 (C-2), 83.9 (C-3), 71.1 (C-4), 76.9 (C-5), 177.9 (C-6); rhamnopyranosyl 102.6 (C-1), 72.0 (C-2), 72.3 (C-3), 73.8 (C-4), 69.5 (C-5), 19.3 (C-6); glucopyranosyl 99.7 (C-1), 79.2 (C-2), 78.3 (C-3), 71.4 (C-4), 78.0 (C-5), 64.0 (C-6); xylopyranosyl 98.7 (C-1), 74.07 (C-2), 75.9 (C-3), 71.8 (C-4), 65.9 (C-5).

ACIDIC HYDROLYSIS OF COMPOUND 1.—Compound **1** (25 mg) was hydrolyzed with 2 M HCl-dioxane (1:1) (3 h) on a boiling H₂O bath to afford the aglycone, crystallized from MeOH (5 ml) as colorless needles: mp 310–315°, identified as oleanolic acid by direct comparison (mmmp, cotlc, and co-ir) with an authentic sample: eims [M]⁺ 456, 441, 411, 248, 208, 192.

The neutralized (Ag₂CO₃) and concentrated hydrolysate showed the presence of D-glucuronic acid and L-rhamnose (pc, solvent D, R_f values 0.15 and 0.37, respectively). The estimation of sugars was performed using colorimetric methods (7).

METHYLATION.—Compound **1** (250 mg) was dissolved in MeOH (5 ml) and was treated with excess ethereal CH₂N₂. The whole reaction mixture was allowed to stand for 12 h. Removal of the solvent under reduced pressure gave **1** methyl ester (220 mg).

NaBH₄ REDUCTION OF 1 METHYL ESTER.—The methyl ester of **1** (100 mg) was dissolved in dry Et₂O (4 ml). NaBH₄ (500 mg) was added, and the mixture was ice-cooled, stirred for 20 h at room temperature, and evaporated in vacuo. The residue was diluted with an excess of H₂O and extracted with *n*-BuOH. The *n*-BuOH layer was washed, dried, and concentrated. It afforded a reduced glycoside of **1** (75 mg), ir max (Csl) cm⁻¹ 3400 (OH), no COOMe.

PERMETHYLATION OF THE REDUCED GLYCOSIDE OF 1.—A solution of the reduced glycoside of **1** (200 mg) in MeOH was treated with dry DMSO (0.4 ml), dry *t*-BuONa (40 mg), finely powdered dry NaOH (10 mg), and MeI (0.3 ml). The reaction mixture was stirred at room temperature for 1 h. The solution was poured into ice-H₂O and extracted with Et₂O. The Et₂O layer was washed with a saturated NaCl solution, dried, and evaporated. The product obtained was purified by cc (SiO₂, solvent C) to furnish the methylated reduced glycoside of **1** (190 mg).

METHANOLYSIS FOLLOWED BY HYDROLYSIS OF THE METHYLATED REDUCED GLYCOSIDE OF 1.—A solution of the methylated reduced glycoside (50 mg) in 7% HCl/dry MeOH (1 ml) was treated under reflux for 3 h and acid-hydrolyzed with 6% H₂SO₄. The reaction mixture was neutralized with Ag₂CO₃ powder and filtered. The mother liquor furnished 2,4,6-tri-*O*-methyl-D-glucose and 2,3,4-tri-*O*-methyl-L-rhamnose (R_G values 0.76 and 1.01, respectively) which were identified with authentic samples on pc (solvent E).

ACIDIC HYDROLYSIS OF COMPOUNDS 2 AND 3.—Compounds 2 and 3 (25 mg each) were separately hydrolyzed with 2 M HCl-dioxane (1:1) (5 ml) for 3 h as usual to afford an aglycone identified as oleanolic acid by its mp, co-tlc, and co-ir with an authentic sample.

The neutralized (Ag₂CO₃) aqueous hydrolysates of 2 and 3 on pc (solvent D) showed the presence of D-glucuronic acid, D-glucose, and L-rhamnose (R_f 0.15, 0.18, 0.37, respectively). Compound 3 showed D-xylose (R_f 0.28) also.

ALKALINE HYDROLYSIS OF 2 AND 3.—Solutions of 2 and 3 (250 mg each) in MeOH (5 ml) were separately treated with 5% aqueous K₂CO₃ (5 ml), and the whole mixture was heated under reflux for 3 h. The reaction mixture was neutralized with Dowex 50W × 8(H⁺) and filtered. Workup of the filtrate in the usual manner yielded a prosapogenol (210 mg), crystallized from MeOH as a colorless crystals. The prosapogenol was found to be identical to compound 1 (mmp, co-tlc, and co-ir).

METHYLATION OF COMPOUNDS 2 AND 3.—Compounds 2 and 3 (100 mg each) were separately methylated with CH₂N₂ as described for compound 1 to furnish their methyl esters (80 and 85 mg, respectively).

NaBH₄ REDUCTION OF THE METHYLESTERS OF 2 AND 3.—The methyl esters of compounds 2 and 3 (60 mg each) were treated with dry Et₂O and NaBH₄ (500 mg) as above to afford the respective methylated reduced glycosides.

PERMETHYLATION FOLLOWED BY HYDROLYSIS OF THE METHYLATED REDUCED GLYCOSIDES OF 2 AND 3.—The methylated reduced glycosides of 2 and 3 (50 mg each) were separately permethylated as above. The permethyl ethers thus obtained were hydrolyzed as above to furnish oleanolic acid. The mother liquors afforded 2,4,6-tri-*O*-methyl-D-glucose, 2,3,4-tri-*O*-methyl-L-rhamnose, and 2,3,4,6-tetra-*O*-methyl-D-glucose from the permethyl ether of reduced 2 and 2,4,6-tri-*O*-methyl-D-glucose, 2,3,4-tri-*O*-methyl-L-rhamnose, 2,3,4-tri-*O*-methyl-D-

xylose, and 3,4,6-tri-*O*-methyl-D-glucose (Wallenfels positive) from the permethyl ether of reduced 3.

PARTIAL HYDROLYSIS OF COMPOUND 3.—Compound 3 (200 mg) in 1 M HCl-*n*-BuOH (1:1) (50 ml) was heated at 70° for 4 h. The *n*-BuOH layer was washed with H₂O and evaporated to dryness in vacuo. The residue after cc (SiO₂, 15 g, solvent C) yielded four prosapogenols tentatively named as PS₁ (40 mg), PS₂ (45 mg), PS₃ (45 mg), and PS₄ (35 mg), in addition to oleanolic acid (20 mg).

PS₂ and PS₃ were found identical to compounds 1 and 2, respectively (mmp and co-tlc).

HYDROLYSIS OF PS₁.—On acidic hydrolysis with 3% H₂SO₄, PS₁ (10 mg) afforded oleanolic acid and D-glucuronic acid and D-glucose. PS₁ on alkaline hydrolysis gave D-glucose.

NaBH₄ REDUCTION AND PERMETHYLATION OF PS₁.—PS₁ was treated with NaBH₄ as above. Permethylation followed by hydrolysis afforded 2,3,4,6-tetra-*O*-methyl-D-glucose (R_G 1.00).

ACIDIC HYDROLYSIS OF PS₄.—PS₄ (10 mg) on acidic hydrolysis afforded oleanolic acid, D-glucose, and D-xylose.

PERMETHYLATION OF PS₄.—PS₄ (20 mg) was permethylated as above, followed by acid hydrolysis, and afforded 2,3,4-tri-*O*-methyl-D-xylose and 3,4,6-tri-*O*-methyl-D-glucose (pc, solvent D, R_G 0.98 and 0.76, respectively).

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